

## HANDBOOK

# Bio-T kit<sup>®</sup> IBDV

Cat. N° BIOTK081 - 100 reactions

**Detection of Infectious Bursal Disease Virus (IBDV), the causative agent of Gumboro disease  
by real-time RT-PCR (qRT-PCR)  
with endogenous (IPC) and exogenous (IC) internal positive control**

## AVIAN

### Sample type

- Bursa of Fabricius swabs;
- Organs or organs homogenates (Bursa of Fabricius, spleen)
- FTA<sup>®</sup> Cards
- Individual analysis or by pool up to 5

### Recommended Nucleic Acids Extraction-Purification

- Silica membrane columns (BioSellal – BioExtract<sup>®</sup> Column Cat. N° BEC050 or BEC250)
- Magnetic beads (BioSellal – BioExtract<sup>®</sup> SuperBall<sup>®</sup> Cat. N° BES384)

*Veterinary use only*



## DOCUMENTS MANAGEMENT

The Bio-T kit® IBDV has two technical handbooks:

- The extraction handbook for Bio-T kit® IBDV displaying BioSella's suggested extraction protocols.
- The Bio-T kit® IBDV qRT-PCR handbook, presenting the instruction information to perform the qRT-PCR.

The latest versions in use for each handbook are indicated on the certificate of analysis (CA) provided with Bio-T kit® IBDV.

Besides these two handbooks, a summary report of the validation file is available on request, contact BioSella (contact@biosellal.com).

## REVISION MANAGEMENT

BioSella indicates modifications done to this document by highlighting them using the rules presented in the Table below:

MODIFICATIONS MANAGEMENT			
Type of modification Highlighting color	Minor modifications	Type 1 Major modifications	Type 2 Major modifications
Impact on revision / version	Change of revision date No change of version	Change of revision date + change of version	Change of revision date + change of version
Examples of modifications	Corrections: typographical, grammatical or turns of phrase	EPC reference modification	Modification of Master Mix composition
	Addition of new sample type for extraction	Exogenous IPC reference modification	Modification of validated extraction protocol
	Addition of information giving more details or alternative protocol		
	Addition/Suppression of optional information		

## PRESENTATION

### Recommendations for sampling, shipping and storage of samples

Real-time RT-PCR is a powerful technique allowing the detection of few amounts of pathogen genome. Genome can be rapidly degraded depending on the pathogen nature (bacteria / parasites, enveloped viruses...), the genome nature (DNA / RNA) and the sample type (presence of DNase / RNase). Thus, BioSellaal recommends the following instructions to guarantee an optimal diagnosis.

#### Sampling

To prevent cross-contamination between samples leading to false positive results, it is mandatory to use disposable materials and to avoid direct contact between specimens.

#### Shipping

It is recommended to ship soon as possible after sampling, under cover of positive cold.

#### Storage after reception

Recommended storage of samples at  $5^{\circ}\text{C} \pm 3$  for a maximum of 7 days and  $\leq -16^{\circ}\text{C}$  beyond.

## AVIAN Line

This kit belongs to the AVIAN Line of BioSellaal which groups together a set of kits that share common extraction and RT-PCR protocols (More information on [www.biosellal.com](http://www.biosellal.com)). It is compatible with BioSellaal's other kits belonging to the PIG line. (information available on [www.biosellal.com](http://www.biosellal.com)).

### Description of Bio-T kit® IBDV

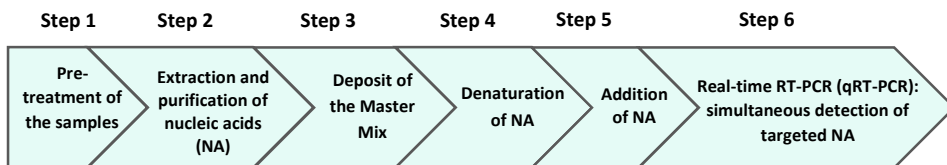
**The Bio-T kit® IBDV** (Cat. N°BIOTK081) contains a ready to use **one-step RT-PCR Master Mix** allowing the detection **in the same reaction well of:**

- **Infectious Bursal Disease Virus (IBDV)** (6-FAM labelled),
- **an exogenous internal positive control RNA (IC)** (VIC labelled) to assess the absence of RT-PCR inhibitors
- **a RNA endogenous internal positive control (IPC)** (beta actin), (Cy5 labelled), to assess the presence of sufficient amount of host cells, sample integrity, nucleic acids extraction quality and absence of RT- PCR inhibitors

This kit can be used for the qualitative analysis of IBDV (detected or not detected) on samples such as swabs and organs. It was developed and validated according to the **French standard NF U47-600-2 published by AFNOR**.

**Extraction protocols suggested by BioSellaal are available on request.**

## Description of the whole process



Pre-processing and extraction protocols on request		qRT-PCR handbook of the Bio-T kit® IBDV			
Organs* FTA® Cards* Organs homogenates* Swabs*	BioExtract® SuperBall®  BioExtract® Column	Ready-to-use Master Mix <b>MMIBDV-A</b>	Denaturation of IBDV dsRNA	Samples NC/NCS EPC (EPCIBDV-A)	Dyes: FAM/VIC/Cy5 Passive reference: ROX Program: PIG/AVIAN program with RT ramping Standard or Fast

\* pre-treatment mandatory

## Kit contents and Storage conditions

**Table 1. Description of the kit contents**

Description	Reference	Volume/tube	Presentation	Conservation
<b>Master Mix (MM)</b> Ready to use	MMIBDV-A	2x750 µl	Tube cap grey Bag A	≤-16°C Protected from light, « MIX » Zone
<b>Exogenous Internal Control (IC)</b>	IPCRNA-A	500 µl	Tube cap purple Bag B	≤-16°C « Nucleic acids Extraction » Zone
<b>External Positive Control (EPC)</b> Positive PCR control of IBDV	EPCIBDV-A	200 µl	Tube cap red Bag C	≤-16°C « Addition of Nucleic acids » Zone
<b>Water</b> RNase/DNase free	Aqua-A	1 ml	Tube cap blue Bag C	5°C ±3 or ≤-16°C « Addition of Nucleic acids » Zone

Kit reagents are stable until the expiration date stated on the label, subject to compliance with good storage conditions.

## List of consumables and reagents not included in the kit

**Table 2. Consumables and reagents not included in the kit**

Consumable / Reagent	Description	Provider	Cat. N°
ATL Buffer	Lysis Buffer	BioSellal	ATL19076
BioExtract® Column	DNA/RNA column extraction kit (50)	BioSellal	BEC050
BioExtract® Column	DNA/RNA column extraction kit (250)	BioSellal	BEC250
BioExtract® SuperBall®	DNA/RNA Magnetic beads extraction kit (4 x 96)	BioSellal	BES384

For consumables related to the thermocycler, refer to the user manual of the device.

## List of reagents to confirm laboratory performance

An IBDV synthetic RNA (titrated in number of copies/RT-PCR), used by BioSellal in his validation file, can be used to confirm the performance of your thermocycler(s). BioSellal sells this reagent under the following reference:

**Table 3. Optional reagent\***

Reagent	Description	Provider	Cat. N°
RNA IBDV	IBDV quantified RNA (7.5 10 <sup>4</sup> copies/qRT-PCR)	BioSellal	cARN-IBDV-001

\*This reagent is available only on demand, please contact BioSellal ([contact@biosellal.com](mailto:contact@biosellal.com)).

## General precautions

- Wear appropriate personal protective equipment (lab coat, disposable gloves frequently changed).
- Work in dedicated and separate areas to avoid contamination: "Extraction" (unextracted samples storage, extraction equipment area), "Mix" (ready to use Master Mix storage, qRT-PCR plates preparation), "Nucleic acids (NA) Addition" (Nucleic Acids storage and addition of extracted NA and controls in the qRT-PCR plate), "PCR" (final area containing the thermocycler(s)).
- Use dedicated equipment for each working area (gloves, lab coat, pipettes, vortex ...).
- Use filter tips.
- Before use, thaw all components at room temperature.
- Vortex and spin briefly (mini-centrifuge) all reagents before use.
- **One-step RT-PCR Master-Mix is less stable than PCR Master-Mix. To guarantee its optimal performance, it is mandatory to defrost the tubes extemporaneously before use, to vortex it just before use, to keep it at 4 ° C during the deposit and to refreeze it immediately afterwards.**
- Avoid the repetition of freezing-thawing cycles for samples, lysates, extracted NA.
- **Working with RNA is more demanding than working with DNA** (RNA instability and omnipresence of the RNases).

For these reasons, special precautions must be taken:

- Always wear gloves, change them frequently, especially after contact with skin or work surfaces.
- Treat all surfaces and equipment with RNases inactivation agents (available commercially).
- When wearing gloves and after material decontamination, minimize the contact with surfaces and equipment in order to avoid the reintroduction of RNases.
- Use "RNase free" consumable.
- It is recommended to store the RNA at  $\leq 5 \pm 3^\circ\text{C}$  during the manipulation and then freeze it as soon as possible, preferably at  $\leq -65^\circ\text{C}$  or by default at  $\leq -16^\circ\text{C}$ .
- Open and close tubes one by one in order to limit the opening times and avoid any contact with RNases present in the environment (skin, dust, working surfaces...).

# DETECTION OF INFECTIOUS BURSAL DISEASE VIRUS BY qRT-PCR WITH BIOTK081

## Global procedure

### 1) Establish qRT-PCR plate setup defining each sample position and including the following controls:

- **Negative Control Sample (NCS):** water (or PBS) replaces the sample from the first step of sample preparation.  
This control is mandatory for each extraction series.
- **Negative Amplification Control (NC):** 5 µl of water RNase/DNase free (Aqua-A tube, **Blue** cap) replaces sample nucleic acids extract on qRT-PCR plate.  
This control is recommended when using the kit for the first time or to verify the absence of Master Mix contamination.
- **IBDV External Positive Control (EPC):** synthetic DNA, containing the targeted sequence specific of IBDV (tube EPCIBDV-A, **red** cap)  
This control is mandatory.

**⚠ CAUTION:** *EPC tube handling represents nucleic acids contamination hazard, it is thus recommended to open and handle it in a restricted area, away from other PCR components and to take precautions to avoid cross-contamination with nucleic acids extracts during deposit on the qRT-PCR plate.*

### 2) Denaturation of nucleic acids (NA)

Double-strand IBDV RNA must be denaturated in single strand before to perform reverse-transcription and PCR.

To this purpose:

- Take 10 to 15 µl of nucleic acids extracts into a microplate – seal the plate
- Incubate during 3 minutes at 95°C±1.5.
- Place **immediately** the nucleic acids for at least 5 minutes at 5 ° C. ± 3 to limit the renaturation of the double-strands and to avoid the introduction into the Master Mix of a solution at high temperature which could degrade the Reverse Transcriptase enzyme.

**It is strongly recommended to perform the dsRNA denaturation step just prior to the preparation of the qRT-PCR plate in order to prevent renaturation of the double stranded RNAs.**

### 3) Preparation of the qRT-PCR plate

#### In the «MIX » dedicated Area

1. After thawing, vortex and rapid centrifugation of the tube, **transfer 15µl of Master Mix MMIBDV-A** (grey cap) in each well of interest (samples and controls).

**⚠ One-step RT-PCR Master-Mix is less stable than PCR Master-Mix. To guarantee its optimal performance, it is mandatory to extemporaneously defrost the tubes just before the use, to vortex it, to keep it at 4 ° C during the deposit and to refreeze it immediately afterwards.**

#### In the «Nucleic Acids addition» dedicated Area

2. **Add 5 µl of nucleic acids extract (or NCS, water, EPC: EPCIBDV-A, red cap)** in each well of interest. Make sure to pipet out the 5 µl in the bottom of the well, in the Master Mix, and to avoid the formation of bubbles.
3. Seal the plate with an optically clear sealer or close the strip caps.

#### In the «PCR» amplification dedicated area

4. **Define the thermocycler parameters** (see Table 4, Table 5, Table 6)
5. It is recommended to **spin down the plate prior to place it into the thermocycler**, in purpose to avoid the presence of drops on the walls of the wells and to eliminate the maximum of bubbles.
6. Start the qRT-PCR program. Approximate duration of the run: 90 minutes.

### 4) Thermocycler settings

This kit was developed and validated on AriaMx™ (ramping Fast) and ABI PRISM® 7500 Fast (ramping Standard or Fast). For other thermocyclers, contact our technical support.

Table 4. Thermocycler Configuration		
	ABI PRISM® 7500 Fast	AriaMx™
Mode	Quantitation – Standard curve	Quantitative PCR, Fluorescence Probe
Ramping	Ramping Standard or Ramping Fast	Ramping Fast
Passive Reference	ROX	ROX

Table 5. Thermocycler settings			
Target	Dyes		Final Volume / well
	Reporter	Quencher	
IBDV	FAM	NFQ-MGB or None*	20 µl  = 15 µl Master Mix + 5 µl nucleic acids or controls†
Exogenous IC	VIC	NFQ-MGB or None*	
Endogenous IPC	CY5	NFQ-MGB or None*	
To assign to samples and controls†			

\* Depending on thermocycler, contact our technical support (tech@biosellal.com)

† Controls are NC (water), NCS and EPC (IBDV).

Table 6. Amplification PROGRAM AVIAN with RT		
Ramping Standard or Fast		
Cycles	Time	Temperature
1 cycle	20 min	50°C
1 cycle	5 min	95°C
40 cycles	10 s	95°C
	45 s	60°C
	+ data acquisition	

NB: Amplification Program are compatible with all kits of AVIAN Lines from BioSellal.

## RESULTS INTERPRETATION

To analyze and interpret the signals obtained by qRT-PCR, the Threshold line must be set up.

The Threshold must be assigned carefully to obtain the most reproducible result between different manipulations according to the requirements defined in Annex C of the French Standard NF U47-600 (part 1). A consistent set of positives curves, at least the EPC, is used to set the threshold value above the baseline and in the exponential amplification phase of the plot.

The Threshold Cycle, named « Ct » or « Cq » (depending on thermocyclers), corresponds to the intersection between the amplification curves and the threshold line. It allows the relative measurement of the concentration of the target in the RT-PCR reaction when a calibrated extract is analyzed in the same qRT-PCR run.

The qRT-PCR run is validated if the controls (EPC, NCS and NC) present valid results, then the result of each sample can be interpreted.



## Main scenarios

### Controls reading

**Table 7. Controls results interpretation**

	Targets			Interpretation
	IBDV (FAM)	Exogenous IC (VIC)	Endogenous IPC (Cy5)	
<b>NCS</b> Negative Control Sample  <b>MANDATORY</b>	Neg	Pos	Neg	Valid
	Pos/ Neg	Pos	Pos/ Neg	If at least one of IPC and IBDV target is positive: Contamination with negative / positive sample during extraction or preparation of plate.
<b>NC</b> Negative Amplification Control  <b>OPTIONNAL</b>	Neg	Neg	Neg	Valid
	At least one target <b>Pos</b>			Contamination with negative / positive sample during extraction or preparation of plate or Master Mix /water contamination.
<b>EPC</b> External Positive Control IBDV  <b>MANDATORY</b>	Pos*	Neg	Neg	Valid
	Neg	Neg	Neg	Master Mix error? EPC omission?
	Pos*	At least one target <b>Pos</b>		Contamination during the preparation of the plate by a sample.

\* Ct value obtained must comply with the value given on the certificate of analysis (CA).

**Note:**

Endogenous IPC targets a gene expressed by avian cells, thus it cannot be detected in NCS, NC and EPC.

## Samples reading: in case of IC addition during extraction or qRT-PCR

**Table 8. Different types of results for samples with IC addition**

Targets			
IBDV (FAM)	Exogenous IC (VIC)	Endogenous IPC (Cy5)	Interpretation
Neg	Pos*	Pos*	Negative or Un-detected
Pos			Positive or Detected
Pos	Pos*	Neg or Ct>35	Positive or Detected Quantity of cells insufficient? Competition with IBDV target?
	Neg or Ct>35	Pos*	Omission of IC addition? Trouble during nucleic acids extraction?
		Neg or Ct>35	RT or PCR inhibitors <sup>†</sup> ?
Neg	At least one valence Neg or Ct>35		<b>Uninterpretable</b> = analysis to be renewed Omission of nucleic acids during plate setup or addition not in contact with the Master Mix during plate preparation? Omission of IC addition? RT or PCR inhibitors <sup>†</sup> ? nucleic acids degradation? Trouble during nucleic acids extraction?

\* Obtained Ct values depends on the thermocycler, the analysed matrix and the extraction methods used. For IC, it must be, at most, within the range specified on the certificate of analysis (CA). IPC and IC values, obtained from the different matrices, are available upon request. BioSella recommends that the laboratory determines its own maximum tolerated IPC value based on its extraction method and thermocycler.

† In case of suspicion of inhibition, 1) Repeat qRT-PCR by pre-diluting nucleic acids to 1/10 or even 1/100 in DNase / RNase free water or 2) Resume analysis from extraction.

## Samples reading: in absence of IC addition during extraction or qRT-PCR

**Table 9. Different types of results for samples without IC addition**

Targets		
IBDV (FAM)	Endogenous IPC (Cy5)	Interpretation
Neg	Pos*	Negative or Un-detected
Pos		Positive or Detected
Pos	Neg or Ct>35	<b>Positive or Detected</b> Quantity of cells insufficient? Competition with IBDV target? Omission of IC addition? Trouble during nucleic acids extraction? RT or PCR inhibitors <sup>†</sup> ?
Neg	Neg or Ct>35	<b>Uninterpretable</b> = analysis to be renewed Omission of nucleic acids during plate setup or addition not in contact with the Master Mix during plate preparation? Omission of IC addition? RT or PCR inhibitors <sup>†</sup> ? nucleic acids degradation? Trouble during nucleic acids extraction?

\* Obtained Ct value depends on the thermocycler, the analysed matrix and the extraction methods used. IPC values, obtained from the different matrices, are available upon request. BioSella recommends that the laboratory determines its own maximum tolerated IPC value based on its extraction method and thermocycler.

† In case of suspicion of inhibition, 1) Repeat qRT-PCR by pre-diluting nucleic acids to 1/10 or even 1/100 in DNase / RNase free water or 2) Resume analysis from extraction.



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